RAPIDStain. For Staining of Polyacrylamide Gels

INTRODUCTION

RAPIDStain. is based on a colloidal Coomassie stain. RAPIDStain. only stains proteins, leaving the background clear,which results in high band visibility. The staining of gels with RAPIDStain. allows the examination of protein bands during the staining process. After the staining process, the band intensity may be further enhanced by equilibration of the stained gel in deionized water. RAPIDStain. has a sensitivity of staining 4-8ng protein/band (e.g., 4-8ng of BSA is visible in 4-20% SDS acrylamide gels).

Catalog No.		Size	Specification
RAPIDStain.	786-31	1L	Suitable for 40-50 Mini gels (8 x 10cm) & 20-25large gels (12 x 15cm)

Storage Condition:

Shipped at ambient temperature. Store at room temperature upon arrival. Stable for one year when stored and used properly.

Instructions For Staining of Polyacrylamide Gels:

1. Wash gel 3 times, 5 minutes each, in deionized water to remove SDS present in the gel. Each wash should be in large volumes of water. For gels without SDS, a single wash in deionized water is sufficient. (Note: Isoelectric focusing gels require prefixing in 20% trichloroacetic acid for 30 minutes, followed by extensive washing to remove TCA).

2. Remove all free water from the gel. Add an adequate amount of RAPIDStain. to cover the gel. Gently shake the gel in stain for 1 hour. Protein bands will be visible within 2-5 minutes and reach a maximum intensity within 1hour. Longer incubation may be performed and will not increase the background.

3. Rinse the stained gel in a large volume of deionized water, 2-3 times for 10-15 minutes each. Rinsing in deionized water enhances the intensity of the protein bands. Store the stained gel in deionized water.

NOTE: If background staining is noticed, it is indicative of residual SDS in the gel. Rinsing the gel extensively in deionized water will remove the background staining.

founded : 2000

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